

## 6- AND 1- SUBSTITUTED MANNOSYL PHOSPHOTRIESTERS AS LIPOPHILIC MACROPHAGE-TARGETED CARRIERS OF ANTIVIRAL NUCLEOSIDES

C. Gouyette<sup>(a)</sup>, J.M. Neumann<sup>(b)</sup>, R. Fauve<sup>(c)</sup>, and T. Huynh-Dinh<sup>(a)\*</sup>

(a) Unité de Chimie Organique, URA CNRS 487, Institut Pasteur, 75724 Paris Cedex 15, France

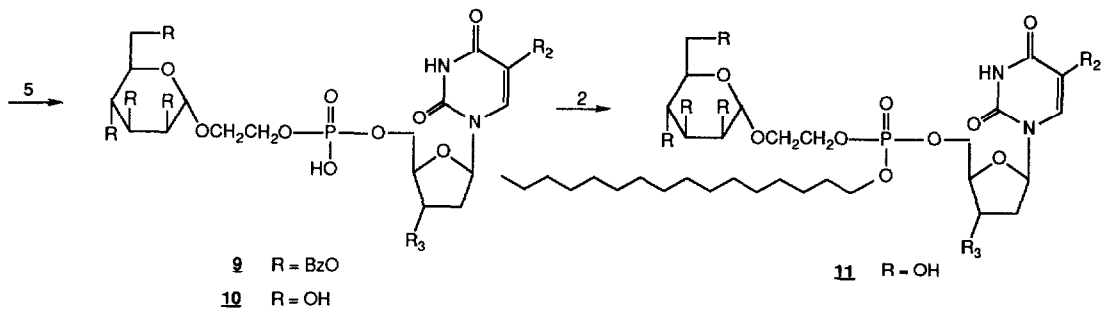
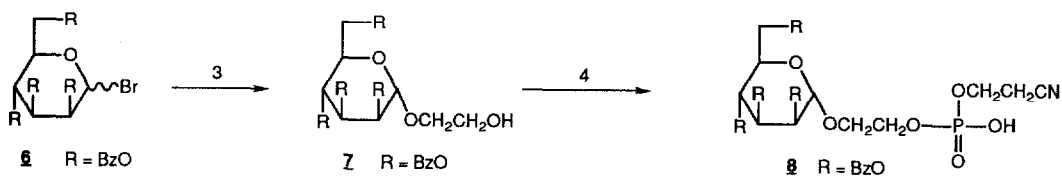
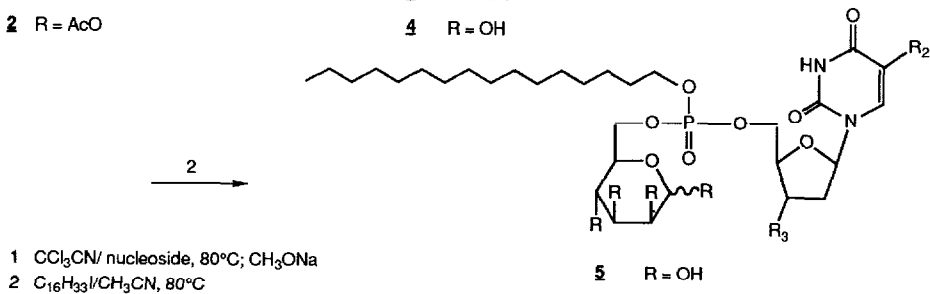
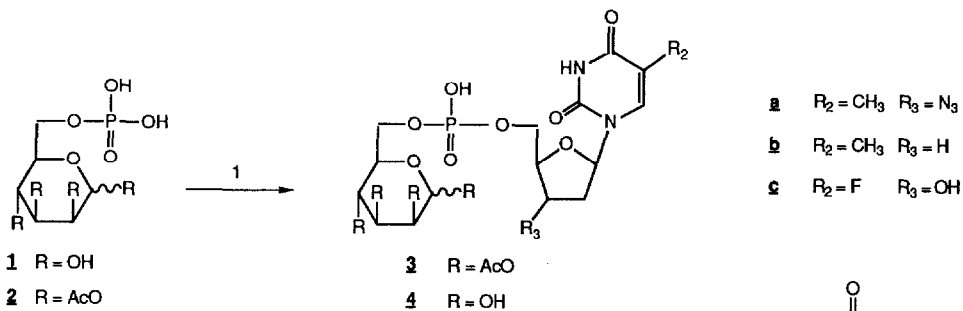
(b) Service de Biophysique, CEN de Saclay, 91191 Gif-sur-Yvette Cedex, France

(c) Unité d'Immunophysiologie Cellulaire, URA CNRS 1113, Institut Pasteur

**Abstract :** Lipophilic mannose-6 phosphate and (1- $\alpha$  mannosyl)ethyl phosphate derivatives of the antiviral nucleosides 3'-azido thymidine (AZT), 2',3'-dideoxy thymidine (ddT) and 2'-deoxy 5-fluoro uridine (dU<sup>F</sup>) were prepared from D-mannose as membrane soluble prodrugs directed towards cells carrying mannosyl receptors.

Starting from the model dolichyl glucose-1 phosphate which serves as a lipid carrier in glycoprotein, we have recently described the synthesis of a hexadecyl 6-D-glucopyranosyl 5'-thymidyl phosphate<sup>1</sup> and its NMR studies of trans-membrane transport across unilamellar vesicles<sup>2</sup>. A series of antiviral nucleosides derived from thymidine have been prepared and their promising biological activities have increased our interest to another carbohydrate series, that of D-mannose. We wondered whether the mannosyl residue, in addition to its higher water solubility, could also be used as a site-directing moiety towards mannosyl-binding proteins. This ligand-cell recognition is important in the case of macrophages, which are known to be the target of numerous pathogens which replicate in these cells and to present mannose-specific receptors responsible for the pathogens adhesion<sup>3</sup>. Moreover, certain HIV strains, proposed as the common factor of AIDS diseases, can replicate in monocytes/macrophages for a considerable time without inducing a substantial cytopathic effect<sup>4</sup> and it was suggested that these cells may be the reservoirs of the virus *in vivo*<sup>5</sup>. It is therefore hoped that compounds which have a deleterious effect on hematopoiesis will not act on bone marrow cells since most of mannosylated drugs will be endocytosed by liver and spleen macrophages; in these conditions a higher therapeutic index could be obtained, together with lower side effects and toxicity.

We wish to report here the synthesis of phosphotriesters of nucleosides, of a lipidic chain and of mannose at C-6 or at C-1 through a C<sub>2</sub> linker, as lipophilic macrophage-targeted carriers of antiviral nucleosides (AZT,



**3** C<sub>6</sub>H<sub>5</sub>CH<sub>2</sub>OCH<sub>2</sub>CH<sub>2</sub>OH/ Hg(CN)<sub>2</sub>/CH<sub>3</sub>NO<sub>2</sub>; Pd-C/ H<sub>2</sub>  
**4** OP(OH)<sub>2</sub>OCH<sub>2</sub>CH<sub>2</sub>CN/ CCl<sub>3</sub>CN, 80°C  
**5** pyridine/ TPSNT/ nucleoside; CH<sub>3</sub>ONa

**a** R<sub>2</sub> = CH<sub>3</sub> R<sub>3</sub> = N<sub>3</sub>  
**b** R<sub>2</sub> = CH<sub>3</sub> R<sub>3</sub> = H  
**c** R<sub>2</sub> = F R<sub>3</sub> = OH

ddT, dU<sup>F</sup>). A similar concept of phosphotriester derivatives of nucleosides analogues was recently presented, associating an alkoxy chain with an aminoacid<sup>6</sup>.

As in the glucose series<sup>1, 2</sup>, the 6-mannosyl phosphotriesters of pyrimidine nucleosides were prepared from D-mannose 6-phosphate **1**: the phosphodiester **3** were obtained by condensation of the protected phosphate **2** and the different nucleosides 3'-azido thymidine (AZT), 2',3' dideoxy-thymidine (ddT) and 2'-deoxy-5-fluoro uridine (dU<sup>F</sup>) with activation of the phosphate function by trichloroacetonitrile. The resulting compounds were deprotected with sodium methylate into **4 a, b, c**, exchanged to their tetrabutyl ammonium salts and alkylated by hexadecyl iodide into the triesters **5 a, b, c** which were isolated after a short-column silica gel chromatography<sup>7</sup>.

We also decided to explore the 1-mannosyl substituted series, which are more closely to the lipidic carrier dolichyl glucose-1 phosphate. As the pyranosyl sugars are converted into 6-phosphates by group-translocation processes during their active transport inside cells, we wanted to obtain glycosyl phosphates where the 6-OH was free in order to be taken up by the phosphotransferase system. To prevent the instability of the phosphate esters at the anomeric position observed in preliminary work, we added a short linker arm to the sugar: 2-benzyloxyethanol reacted with 1-bromo-2,3,4,6-tetrabenzoyl D-mannose<sup>8</sup> **6** to give, after catalytic hydrogenation, 64 % of hydroxyethyl  $\alpha$ -D-mannopyranoside **7**. The mannosyl alcohol **7** reacted with cyanoethyl phosphate in a mixture of CCl<sub>3</sub>CN-pyridine at 80° C overnight to give the phosphate **8** (67 %) which was condensed with the nucleosides into the protected phosphodiester **9 a, b, c** (56-69 %) using triisopropylbenzenesulfonyl-triazole (TPSNT) as an activating agent<sup>9</sup>. Alkaline deprotection gave the phosphodiester **10 a, b, c**. These mannosyl derivatives are closely related to uridine 5'-diphosphate analogs recently prepared by van Boom et al.<sup>10</sup> as inhibitors of glycolipid biosynthesis. The alkylation of the tetrabutylammonium salts of phosphodiester **10** gave, as previously, the mannosyl phosphotriesters **11 a, b, c** (67-70 %).

The purity of all the phosphotriesters **5** and **11** are checked by analytical reverse-phase HPLC and their structures identified by NMR and mass spectra<sup>7</sup>. Preliminary screening have shown antiviral activities for the described compounds; detailed antiviral and NMR trans-membrane transport studies of these 6- and 1-substituted mannosyl phosphotriesters will be reported elsewhere.

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## REFERENCES

- [ 1] IGLESIAS GUERRA F., NEUMANN J.M. and HUYNH-DINH T., *Tetrahedron Lett.*, (1987), **28**, 3581.
- [ 2] NEUMANN J.M., HERVE M., DEBOUZY J.C., IGLESIAS GUERRA F., GOUYETTE C., DUPRAZ B. and HUYNH-DINH T., *J. Am. Chem. Soc.*, (1989), **111**, 4270.

- [ 3] STAHL P.D., WILEMAN T.E. and SHEPHERD V.L., in "Mononuclear Phagocytes - Characteristics, Physiology and Function", VAN FURTH R., *Edit. Nijhoff Publi.*, Dordrecht (1985), 59.
- [ 4] HO D.D., POMERANTZ R.J. and KAPLAN J.C., *N. Engl. J. Med.*, (1987), 317, 278.
- [ 5] GALLO R.C. and MONTAGNIER L., *Sci. Am.*, (1988), 259 (4), 41.
- [ 6] WILLIAMSON J.D. and KINCHINGTON D., Poster Abst. MCP 70, V<sup>th</sup> International Conference on AIDS, Montreal, June 1989.
- [ 7] 5a FAB MS : 734 (MH<sup>+</sup>). NMR (DMSO d<sub>6</sub>) : <sup>1</sup>H chain CH<sub>3</sub> 0.87, (CH<sub>2</sub>)<sub>n</sub> 1.25, CH<sub>2</sub>-CH<sub>2</sub>OP 1.63, CH<sub>2</sub>-CH<sub>2</sub>-OP 4.01; mannose H-1 α 4.90, H-2 3.58, H-3 3.57, H-4 3.45, H-5 3.72, H-6 4.23, H-6' 4.09 ; nucleotide H-6 7.51 (41 %) and 7.49 (59 %). CH<sub>3</sub> 1.82, H-1' 6.14 ; <sup>31</sup>P (TMP) : -3.65 (50 %) and -3.84 (50 %). TLC : 0.67 (isopropanol/NH<sub>4</sub>OH/H<sub>2</sub>O 7/1/2).
- 5b FAB MS : 716 (MNa<sup>+</sup>). NMR (DMSO d<sub>6</sub>) : <sup>1</sup>H chain CH<sub>3</sub> 0.87, (CH<sub>2</sub>)<sub>n</sub> 1.25, CH<sub>2</sub>-CH<sub>2</sub>-OP 1.59, CH<sub>2</sub>-CH<sub>2</sub>-OP 4.00; mannose H-1 α 4.89, H-2 3.57, H-3 3.53, H-4 3.40, H-5 3.71, H-6 4.07, H-6' 4.20; nucleotide H-6 7.53 (45%) and 7.50 (55%) CH<sub>3</sub> 1.81, H-1' 6.02. TLC : 0.63.
- 5c FAB MS : 735 (MNa<sup>+</sup>). NMR (DMSO d<sub>6</sub>) : <sup>1</sup>H chain CH<sub>3</sub> 0.85, (CH<sub>2</sub>)<sub>n</sub> 1.24, CH<sub>2</sub>-CH<sub>2</sub>-OP 1.58, CH<sub>2</sub>-CH<sub>2</sub>-OP 3.99 ; mannose H-1 4.90, H-2 3.57, H-3 3.53, H-4 3.41, H-5 3.71, H-6 4.06, H-6' 4.14 ; nucleotide H-6 7.88 (55 %) and 7.85 (55 %), H-1' 6.15 ; <sup>31</sup>P (TMP) : -3.45 (55 %) and -3.82 (45 %). TLC : 0.63.
- 11a FAB MS : 776 (M<sup>+</sup>). NMR (DMSO d<sub>6</sub>) : <sup>1</sup>H chain CH<sub>3</sub> 0.85, (CH<sub>2</sub>)<sub>n</sub> 1.25, CH<sub>2</sub>-CH<sub>2</sub>-OP 1.59, CH<sub>2</sub>-CH<sub>2</sub>-OP 4.00 ; mannose H-1 4.69, H-2 3.63, H-3 3.48, H-4 3.35, H-5 3.42, H-6 and H-6' 3.50, CH<sub>2</sub>-CH<sub>2</sub>-OP 3.59 and 3.79, CH<sub>2</sub>-CH<sub>2</sub>-OP 4.18 ; nucleotide H-6 7.46, CH<sub>3</sub> 1.80, H-1' 6.13 ; <sup>31</sup>P (TMP) : -3.65 (52 %) and -3.82 (48 %); TLC 0.69.
- 11b FAB MS : 759 (MNa<sup>+</sup>) ; NMR (DMSO d<sub>6</sub>) : <sup>1</sup>H chain CH<sub>3</sub> 0.87, (CH<sub>2</sub>)<sub>n</sub> 1.25, CH<sub>2</sub>-CH<sub>2</sub>-OP 1.59, CH<sub>2</sub>-CH<sub>2</sub>-OP 4.00 ; mannose H-1 α 4.67, H-2 3.61, H-3 to H-6, H-6' 3.4 ± 0.1; CH<sub>2</sub>-CH<sub>2</sub>-OP 3.60 and 3.75; CH<sub>2</sub>-CH<sub>2</sub>-OP 4.13 ; nucleotide H-6 7.53 (52%) and 7.51 (48%), CH<sub>3</sub> 1.81, H-1' 6.02. TLC 0.69.
- 11c FAB MS : 780 (MNa<sup>+</sup>). NMR (DMSO d<sub>6</sub>) : <sup>1</sup>H chain CH<sub>3</sub> 0.89, (CH<sub>2</sub>)<sub>n</sub> 1.25, CH<sub>2</sub>-CH<sub>2</sub>-OP 1.60, CH<sub>2</sub>-CH<sub>2</sub>-OP 3.95 ; mannose H-1 α 4.67, H-2 3.64, H-3 3.46, H-4 3.32, H-5-H-6, H-6' 3.45 -<sup>+</sup> 0.05 CH<sub>2</sub>-CH<sub>2</sub>-OP 3.61 and 3.77 ; CH<sub>2</sub>-CH<sub>2</sub>-OP 4.11 ; nucleotide H-6 7.90 (45 %) and 7.88 (55 %), H-1' 6.17 ; <sup>31</sup>P (TMP) : -3.57 (48 %) and -3.90 (52 %). TLC : 0.67.
- [ 8] NESS R.K., FLETCHER H.G. Jr and HUDSON C.S., *J. Am. Chem. Soc.*, (1950), 72, 2200.
- [ 9] "Oligonucleotide Synthesis", GAIT M.J., *Edit. IRL Press*, Oxford (1984).
- [10] BROXTERMAN H.J.G., VAN DER MAREL G.A. and VAN BOOM J.H., *Tetrahedron Lett.*, (1988), 29, 4893.

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